

Decreased VDR Expression in Cutaneous Melanomas as Marker of Tumor Progression: New Data and Analyses

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Abstract. *Background:* Vitamin D₃, acting via vitamin D receptor (VDR) affects a wide range of biological activities, including inhibition of proliferation and angiogenesis, with net antitumor effects. VDR expression is disturbed in many tumors, including melanomas. *Aim:* To find correlation between VDR expression in melanomas and prognostic biomarkers. *Materials and Methods:* VDR was analyzed immunohistochemically in 69 cutaneous melanomas in relation to prognostic factors. *Results:* Less advanced melanomas showed significantly higher VDR expression than the advanced stages. The presence of other markers such as ulceration and lack or non-brisk tumor infiltrating lymphocytes (TILs) was accompanied by significantly lower VDR expression. VDR expression also affected overall survival (OS) with most noticeable effect in the cases without ulceration. *Conclusion:* High VDR expression determines a less malignant phenotype and is related to better prognosis. Loss of VDR expression affects melanoma tumor behavior, allowing for progression of disease. VDR expression can also serve as a prognostic marker in routine histopathology evaluation.

Ultraviolet radiation (UVR) is considered as a one of the most important environmental factors responsible for development of melanoma, an aggressive skin tumor with a relatively high

mortality rate (1). Melanoma originates from melanocytes, cells responsible for production of melanin that protect skin against the damaging effect of UVR (2, 3). UVB photoconverts cutaneous 7-dehydrocholesterol into vitamin D, which is further hydroxylated in the liver at position 25 [by D-25-hydroxylase (CYP27A1)] and in the kidney at positions 1 [by 25-hydroxyvitamin D-1 α -hydroxylase (CYP27B1)] to the biologically-active form 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃; calcitriol) (4, 5). Calcitriol is also produced locally in a number of tissues, including skin (4, 5), where it is involved in regulation of cell proliferation, differentiation or apoptosis in autocrine or paracrine manners (5-7). Alternative pathways of vitamin D activation to biologically active novel vitamin D derivatives were also described recently (8-11).

Calcitriol, in addition to regulation of calcium-phosphate homeostasis, has wide spectrum of pleiotropic effects such as regulation of the adaptive and innate immune activity, endocrine activities and acts as a tumorostatic factor and protector against oxidative stress (4, 12). It also inhibits angiogenesis and regulates expression of adhesion molecules (13, 14). These effects are mediated through an interaction with vitamin D receptor (VDR) that is ubiquitously expressed in the body, including by melanoma cells (5, 15). However, alternative nuclear receptors for vitamin D derivatives were also identified (16). Novel vitamin D₃ analogs that are non-calcemic also have similar antitumor and anti-melanoma activities (17-21).

Our previous studies showed an inverse correlation between VDR expression and progression of skin pigmented lesions (22). Similar correlation was observed for melanomas stratified according Breslow's thickness and Clarks's level, with a decrease of VDR expression in advanced lesions (22). VDR expression affected survival of patients with melanoma (22). Lack of or reduced VDR expression in melanomas localized to the skin lesions or metastases was related to shorter overall survival. The level of VDR was reduced in strongly-melanized melanomas in comparison to amelanotic or slightly-pigmented melanomas (22). A high level of

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melanoma melanization was related to shorter overall survival and disease-free survival (DFS) in metastasizing melanomas (23). Our subsequent research (24) revealed that, similarly to VDR immunostaining, expression of CYP27B1 was inversely correlated with melanoma progression, marked by significantly reduced CYP27B1 expression in most advanced tumors (24). There was no correlation between CYP27B1 expression and ulceration or presence of tumor infiltrating lymphocytes (TILs). However, ulceration and lack of, or a low rate of, TILs were more frequent in melanomas with low or no expression of CYP27B. CYP27B1 expression also affected prognosis, *e.g.* its lack or a low level was related to significantly shorter overall and disease-free survival (24).

In the present study, we analyzed VDR expression in the same cohort of patients with melanoma (22) in relation to newly-collected pathomorphological data as follows: the American Joint Committee on Cancer (AJCC) pTNM classification, overall stage, and other prognostic markers such as ulceration and presence of TILs, to verify the involvement of VDR in melanoma behavior and its potential role as a prognostic and predictive marker.

Materials and Methods

Patients and pathomorphological assessment. The study was approved by the Committee of Ethics of Scientific Research of Collegium Medicum of Nicolaus Copernicus University, Poland (approval number KB 448/2009). Samples of melanomas were obtained from 69 patients, who underwent surgical excision of skin lesions in the Oncology Centre, Prof. Franciszek Łukaszczyk Memorial Hospital, Bydgoszcz, Poland, from 2003 to 2009. Clinicopathological data were obtained from database of the Oncology Center and date of deaths from Department of Registry Office in Bydgoszcz, Poland, and have been updated for 24 months from the end of experiments. The analyzed cohort comprised of 30 superficial-spreading, 37 nodular and 2 acral lentiginous melanomas. Standard hematoxylin and eosin-stained sections of melanomas included in this study were assessed, classified and described by a board-certified pathologist (WJ). The following features were evaluated: 2009 AJCC pTNM classification and stage, ulceration and TILs. The term TILs was used for lymphocytes disrupting or surrounding tumor cells of the vertical growth phase (VGP) (25, 26). Brisk TILs corresponded to lymphocytes infiltrating the entire invasive component of melanoma or the entire base of the VGP. Non-brisk TILs represented only focal lymphocytic infiltrate of the tumor or lymphocytes infiltrating less than the entire base of the VGP. The term absent TILs was used when lymphocytes were absent or were present but did not infiltrate the tumor. The clinicopathomorphological characteristics of melanomas are presented in Table I.

VDR immunohistochemical staining protocol and evaluation. VDR expression was assayed using immunohistochemistry as previously described (22, 27). Briefly, standard formalin-fixed, paraffin-embedded 4- μ m sections were stained overnight using rat antibody to VDR at a dilution of 1:75 (clone 97A; Abcam Inc.,

Table I. *Clinicopathological characteristics of patients with melanoma.*

Features	No. of patients (%)
Age, years	
<25	1 (1.5%)
26-35	3 (4.4%)
36-45	6 (8.7%)
46-55	19 (27.5%)
56-65	17 (24.6%)
66-75	13 (18.8%)
>75	10 (14.5%)
pT	
pT1	15 (27.7%)
pT2	10 (14.5%)
pT3	15 (27.7%)
pT4	29 (42.0%)
pN*	
pN0	34 (49.3%)
pN1	15 (21.7%)
pN2	11 (15.9%)
pN3	8 (11.6%)
pM	
pM0	64 (92.6%)
pM1	5 (7.6%)
Overall stage	
1	17 (24.6%)
2	17 (24.6%)
3	28 (40.6%)
4	7 (10.1%)
Ulceration	
Absent	31 (44.9%)
Present	38 (55.1%)
TILs	
Absent	16 (23.1%)
Non-brisk	32 (46.4%)
Brisk	21 (30.4%)

*There is lack of pN status for one patient.

Cambridge, MA, USA), followed by incubation with anti-rat secondary antibody, conjugated with alkaline phosphatase and red alkaline phosphatase substrate (Vector Laboratories Inc., Burlingame, CA, USA).

Immunohistochemical evaluation was performed by two observers (WJ and AAB) in a blind manner, without knowing the detailed histopathological diagnosis, the newly-evaluated features mentioned above and the other clinical data. The intensity of VDR immunostaining of melanomas was scored semiquantitatively as previously described. Briefly, the staining intensity was scored from 0 to 3 arbitrary units (AU), evaluated in relation to VDR immunostaining in epidermis of normal skin. For statistical analysis purposes, moderate and strong VDR immunostaining was classified as high staining intensity, weak as low staining intensity, and negative as no staining intensity.

Statistical analyses. Statistical analysis was performed with the Prism 5.00 (GraphPad Software, San Diego, CA, USA). Results were considered as statistically significant when the *p*-value was less than 0.05. Data are presented as the mean \pm SD.

Table II. Cytoplasmic and nuclear immunolocalization of vitamin D receptor (VDR) in melanomas stratified according to American Joint Committee on Cancer pTNM classification, overall stage, presence or absence of ulceration and of tumor infiltrating lymphocytes (TILs).

	Cytoplasmic VDR immunostaining (n/%)			Nuclear VDR immunostaining (n/%)		
	None	Low	High	None	Low	High
pT1 (n=15)	2/13.3	7/46.7	6/40.0	1/6.7	10/66.7	4/26.7
pT2 (n=10)	2/20.0	7/70.0	1/10.0	2/20.0	5/50.0	3/30.0
pT3 (n=15)	8/53.3	6/40.0	1/6.7	2/13.3	11/73.3	2/13.3
pT4 (n=29)	11/37.9	18/62.1	0/0.0	6/20.7	20/69.0	3/10.3
pN0 (n=34)	11/32.3	15/44.1	8/23.5	7/20.6	18/52.9	9/26.5
pN1 (n=15)	6/40.0	9/60.0	0/0.0	1/6.7	12/80.0	2/13.3
pN2 (n=11)	0/0.0	11/100.0	0/0.0	0/0.0	10/90.9	1/9.1
pN3 (n=8)	5/62.5	3/37.5	0/0.0	2/25.0	6/75.0	0/0.0
M0 (n=64)	20/31.2	36/56.3	8/12.5	12/18.8	43/67.2	9/14.1
M1 (n=5)	3/60.0	2/40.0	0/0.0	2/40.0	3/60.0	0/0.0
Stage 1 (n=17)	2/11.8	8/47.1	7/41.2	1/5.9	10/58.8	6/35.3
Stage 2 (n=17)	9/52.9	7/41.2	1/5.9	6/35.3	8/47.1	3/17.6
Stage 3 (n=28)	8/28.6	20/71.4	0/0.0	1/3.6	24/85.7	3/10.7
Stage 4 (n=7)	4/57.1	3/42.9	0/0.0	3/42.9	4/57.1	0/0.0
Without ulceration (n=31)	6/19.6	17/54.8	8/25.8	4/12.9	19/61.3	8/25.8
With ulceration (n=38)	17/44.7	21/55.3	0/0.0	7/18.4	27/71.1	4/10.5
TILs absent (n=16)	5/31.3	10/62.5	1/6.3	2/12.5	10/62.5	4/25.0
Non-brisk TILs (n=32)	11/34.4	20/62.5	1/3.1	6/18.8	24/75.0	2/6.3
Brisk TILs (n=21)	7/33.3	8/38.1	6/28.6	3/14.3	12/57.1	6/28.6

Results

VDR expression in relation to pTNM AJCC classification of melanomas. As we reported previously (22), VDR expression was localized in both the cytoplasm and nuclei of melanoma cells. Cytoplasmic VDR was observed in 46 cases (66.7%) and nuclear in 58 cases (84.1%). High VDR expression was seen in the cytoplasm of only eight cases (11.6%), and in the nucleus of 12 cases (17.4%). In the majority of melanomas, a low staining intensity of VDR was observed in the cytoplasm and nucleus in 38 cases (55.1%) and 46 cases (66.7%), respectively) (Figure 1). Table II summarizes the association between VDR expression and pTNM features, overall stage, presence of ulceration and TILs of analyzed melanomas.

VDR expression was inversely-correlated with melanoma progression. Less advanced melanomas exhibited higher VDR expression. Melanomas at stages pT1 and pT2 more frequently had higher VDR immunostaining than melanomas at stages pT3 and pT4 (Table II). On the contrary, melanomas without VDR expression were seen predominantly in advanced stages (pT3 and pT4, Table II). Significant differences were observed between less (pT1 and pT2) and more (pT3 and pT4) advanced stages ($p \leq 0.0001$ and $p = 0.0161$ for cytoplasmic and nuclear staining, respectively). These differences were more visible in the cytoplasm (Figure 2A), because the mean VDR expression was significantly higher in pT1 versus pT3 and pT4

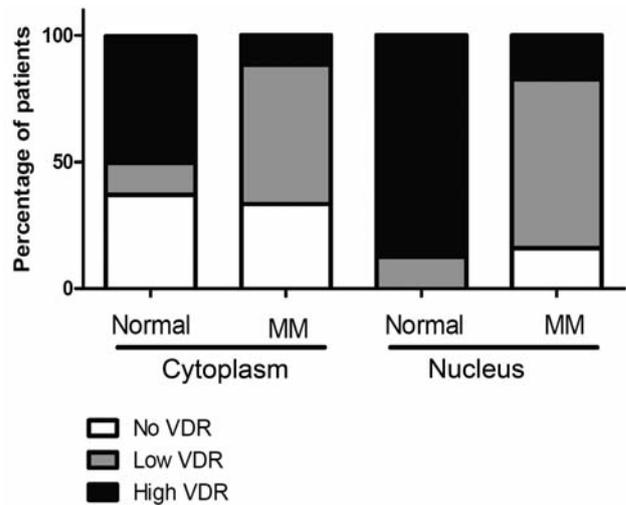


Figure 1. Distribution of cytoplasmic and nuclear vitamin D receptor (VDR) levels in normal skin and malignant melanoma (MM).

(ANOVA) and in pT2 versus pT3 and pT4 (*t*-test). For nuclear VDR immunostaining, the only significant difference observed was for pT1 versus pT4 stages (*t*-test) (Figure 2B).

Melanomas with lymph node metastases exhibited a lack or low VDR expression. No single case at stage pN1-3 exhibited high cytoplasmic VDR immunostaining (Table II). High nuclear VDR staining was observed in two (13.3%) and

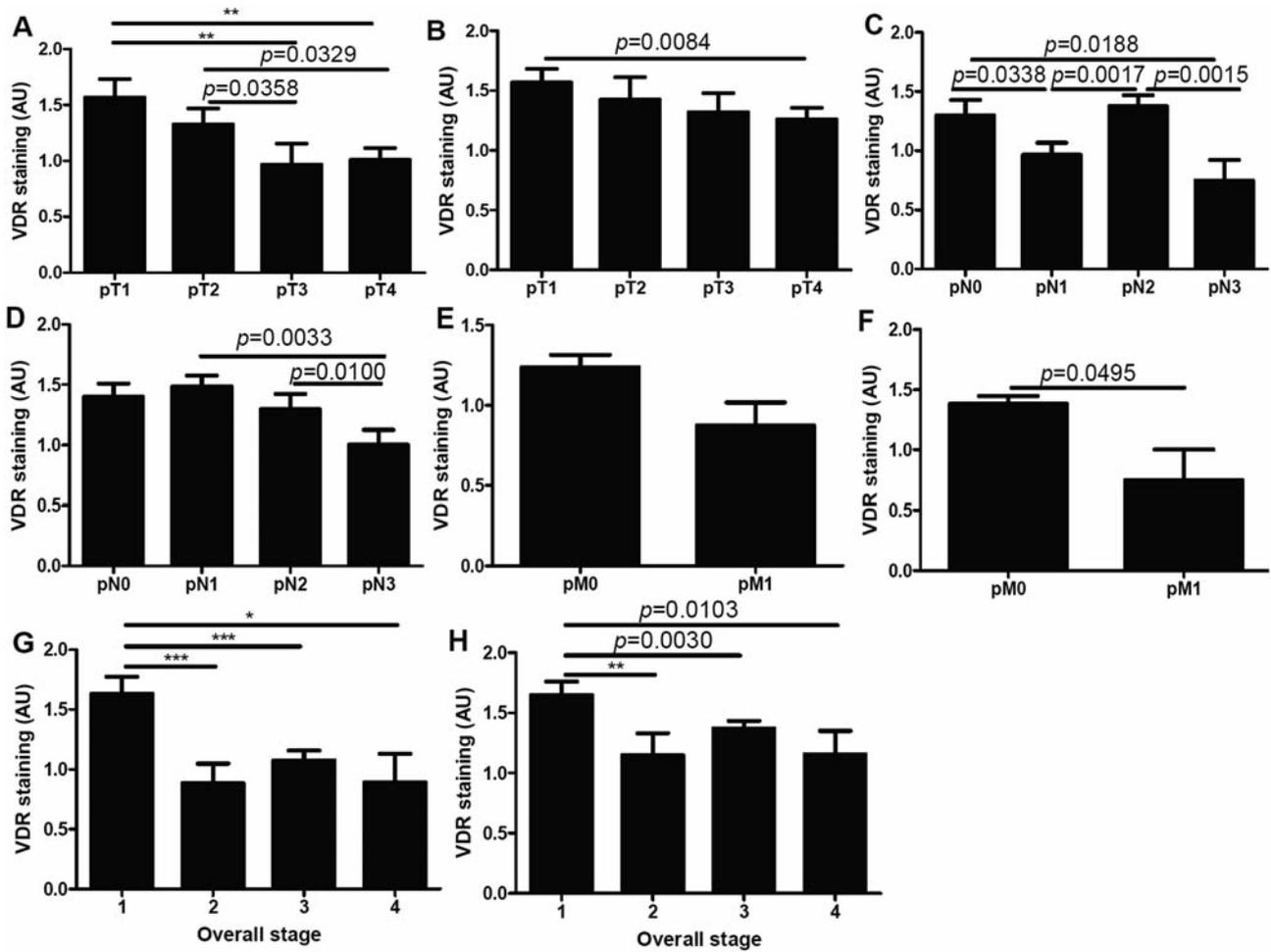


Figure 2. Mean cytoplasmic (A, C, E, G) and nuclear (B, D, F, H) expression of vitamin D receptor (VDR) in melanomas of different pT (A, B), pN (C, D), pM (E, F) and overall (G, H) stage. Statistically significant differences are denoted with p-values as determined by t-test, and with asterisks as * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$ by ANOVA.

one (9.1%) stage pN1 and pN2 cases, respectively. Melanomas at stage pN3 exhibited low or no VDR expression (Table II). In melanomas with four or more lymph node metastases (pN3), nuclear VDR expression was significantly lower than in melanomas without metastases or with less advanced pN stage. This relationship was observed only for nuclear VDR staining (Figure 2 C and D).

The same correlation was seen for pM feature (Figure 2 E and F). In contrast to melanomas at stage pM0, melanomas at stage pM1 exhibited lack of high VDR immunostaining both in the cytoplasm and nucleus. In patients with distant metastases (pM1), a lower mean VDR expression was seen in both the cytoplasm and nucleus when compared to pM0 melanomas. However, these differences were statistically significant only for nuclear immunostaining. It should be noted that only five cases with pM1 stage were included in this study.

Overall melanoma stage was also related to VDR expression. More than 80% of melanomas at a less advanced overall stage (Stage 1) exhibited VDR expression, with 41.2% and 35.3% of cases showing strong cytoplasmic and nuclear immunostaining, respectively. On the contrary, none of the melanomas at stage 4 had high VDR staining (Table II). Melanomas at less advanced stage 1 were characterized by higher mean VDR immunostaining when compared to more advanced melanomas (Stages 2-4). These differences were seen both for cytoplasmic and nuclear VDR expression (Figure 2 G and H).

VDR expression is related to pathological prognostic markers. The relationship between VDR expression and the presence of ulceration was also observed. No single case with high cytoplasmic VDR immunostaining and ulceration was found (Table II). Lack of cytoplasmic VDR expression

was more frequent in melanomas with ulceration: 44.7% of cases of ulcerated melanomas *versus* 19.6% of cases without ulceration (Table II). High nuclear staining was seen in melanomas both with and without ulceration, but the frequency of such staining was less common in the former tumors: 10.5% *versus* 25.8% (Table II). Cases without ulceration had a significantly higher mean cytoplasmic and nuclear VDR expression than ulcerated melanomas (Figure 3 A, B, G and H).

In our population of patients with melanoma, the presence of ulceration was related to a shorter OS [log-rank (Mantel-Cox) test: $\chi^2=7.180$, $p=0.0074$; Gehan-Breslow-Wilcoxon Test: $\chi^2=6.876$, $p=0.0087$] (data not shown).

In melanomas without cytoplasmic VDR, the presence of ulceration had no impact on OS, but in melanomas expressing cytoplasmic VDR the ulceration was related to shorter OS [cytoplasmic VDR and no ulceration *versus* VDR and ulcer: 2297 *versus* 525 days (median) of OS, respectively; log-rank (Mantel-Cox) test: $\chi^2=9.447$, $p=0.0021$; Gehan-Breslow-Wilcoxon test: $\chi^2=8.542$, $p=0.0032$] (Figure 3 C). In melanomas without nuclear VDR, the presence of ulceration had no impact on OS, but in melanomas expressing nuclear VDR, ulceration correlated with a shorter OS [VDR present and no ulceration *versus* VDR present and ulceration: 2297 *versus* 663 days (median) OS, respectively; log-rank (Mantel-Cox) test: $\chi^2=7.158$, $p=0.0075$; Gehan-Breslow-Wilcoxon test: $\chi^2=7.555$, $p=0.0060$] (Figure 3D).

The relationship between lack or low-level of both cytoplasmic and nuclear VDR with shorter survival was more obvious in the subgroup of patients with melanomas without ulceration [log-rank (Mantel-Cox) test: $\chi^2=6.125$, $p=0.0468$ for cytoplasmic VDR staining and log-rank (Mantel-Cox) test: $\chi^2=8.632$, $p=0.0134$; Gehan-Breslow-Wilcoxon test: $\chi^2=6.974$, $p=0.0083$ for nuclear VDR staining] (Figure 3E and F). More detailed analysis demonstrated statistically significant differences between survival curves for patients with lack of VDR *versus* high nuclear VDR immunostaining [log-rank (Mantel-Cox) test $\chi^2=11.250$, $p=0.0008$; Gehan-Breslow-Wilcoxon test: $\chi^2=10.500$, $p=0.0012$] (Figure 3F). Survival curve comparison also showed statistically significant differences for cases with a lack of *versus* low cytoplasmic VDR staining [log-rank (Mantel-Cox) test: $\chi^2=4.564$, $p=0.0327$; Gehan-Breslow-Wilcoxon test: $\chi^2=4.301$, $p=0.0381$] and low *versus* high VDR staining [log-rank (Mantel-Cox) test: $\chi^2=4.695$, $p=0.0302$; Gehan-Breslow-Wilcoxon test: $\chi^2=4.219$, $p=0.0400$] (Figure 3 E).

In patients with melanoma included in this study, the presence of TILs was related to a better prognosis, and OS of patients with TILs was significantly longer when compared to patients with non-brisk TILs [log-rank (Mantel-Cox) test: $\chi^2=8.387$, $p=0.0038$; Gehan-Breslow-Wilcoxon test: $\chi^2=7.181$, $p=0.0074$] and to patients without TILs [log-rank

(Mantel-Cox) test: $\chi^2=6.970$, $p=0.0083$; Gehan-Breslow-Wilcoxon test: $\chi^2=7.266$, $p=0.0070$] (data not shown).

The level of VDR expression was also related to the presence of TILs (Figure 4A-D). In melanomas with brisk TILs, VDR expression was higher, and for cytoplasmic VDR immunostaining, these differences were statistically significant ($p<0.0133$). In melanomas without cytoplasmic VDR, there was a lack of correlation between brisk TILs and OS. However, in melanomas with cytoplasmic VDR, the presence of brisk TILs correlated with significantly better prognosis (median OS: 2297 days for melanomas with brisk TILs *versus* 715 days for melanomas without or with non-brisk TILs, log-rank (Mantel-Cox) test: $\chi^2=8.809$, $p=0.0030$; Gehan-Breslow-Wilcoxon test: $\chi^2=7.536$, $p=0.0060$). A similar trend was observed for nuclear VDR (median OS: 2520 days for melanomas with brisk TILs *versus* 748 days for melanomas without or with non-brisk TILs, log-rank (Mantel-Cox) test: $\chi^2=9.521$, $p=0.0020$; Gehan-Breslow-Wilcoxon test: $\chi^2=7.835$, $p=0.0051$) (data not shown).

There was no relationship between melanin level and the presence of ulceration or TILs. However, a slight decrease of melanin level in melanomas with brisk TILs was observed (data not shown).

Discussion

In the present study, we found significant associations between VDR expression, melanoma progression and prognostic factors in melanoma. In melanomas at lower stages (pT1-2, overall stage 1), statistically significant higher VDR expression was observed. Similarly, melanomas with fewer than three lymph node metastases and melanomas without distant metastases had the strongest VDR expression. Moreover, ulcerated melanomas without TILs or with non-brisk TILs were characterized by lower VDR expression. Furthermore, survival of patients in sub-groups of melanomas without ulcerations was affected by VDR expression, with better prognosis in patients with melanomas expressing VDR. These results are consistent with our previous studies demonstrating decreased expression of VDR during progression of pigmented lesions, including melanoma progression (22). Analysis of OS and well-established factors of unfavorable prognosis showed that the presence of ulceration and lack of TILs or presence of non-brisk TILs correlated with significantly shorter OS.

These results are also consistent with experiments showing inhibition of melanoma proliferation in cell culture by vitamin D₃ (21, 28), which was seen only in cells expressing VDR (29). Similar anti-melanoma activity was shown for other active vitamin D₃ derivatives in human and rodent melanomas grown in monolayer or soft agar (20, 21, 27). In addition, 1,25(OH)₂D₃ inhibited metastases of B16 melanoma in mice (30).

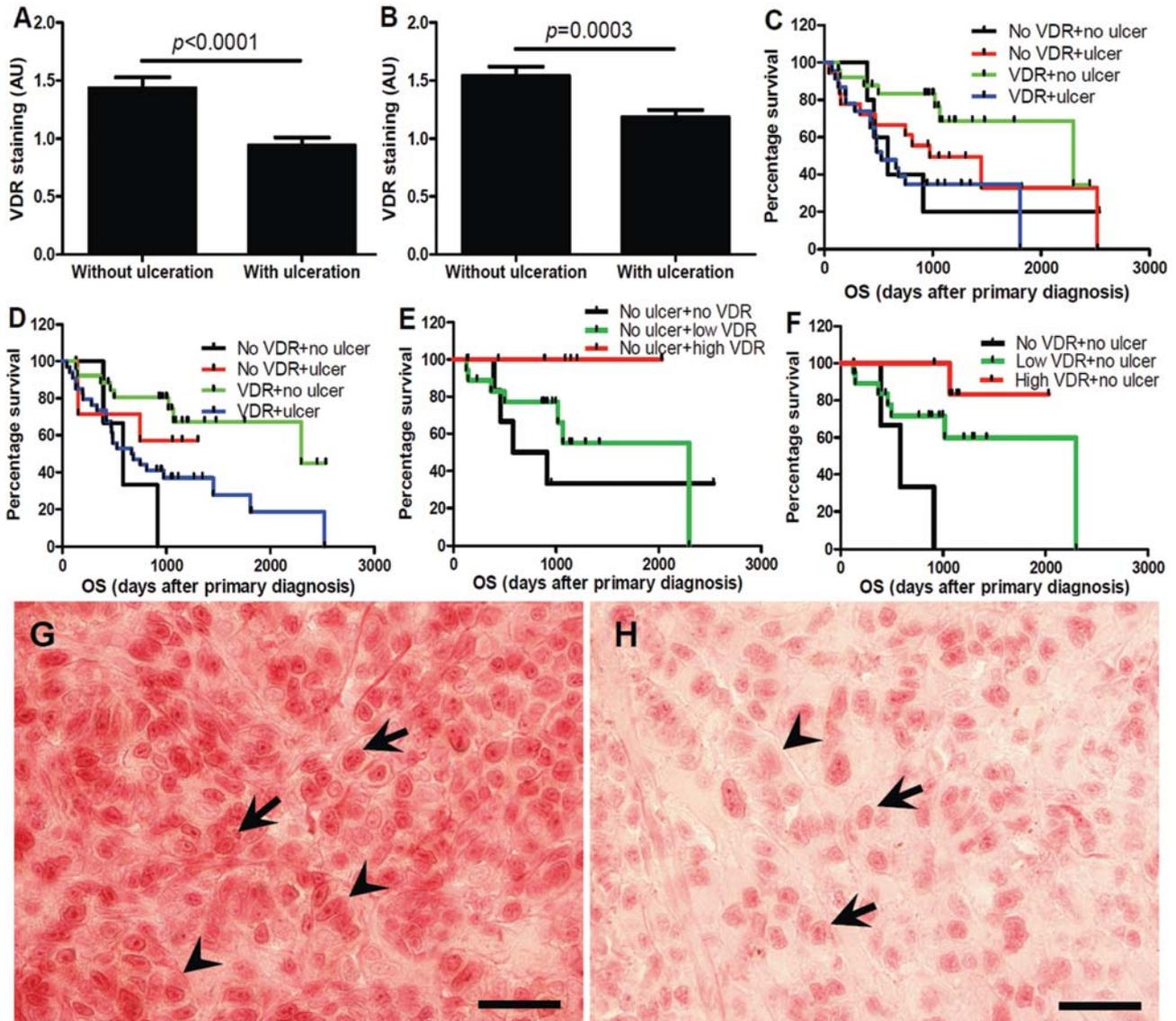


Figure 3. Mean cytoplasmic (A) and nuclear (B) expression of vitamin D receptor (VDR) in melanomas with and without ulceration. Statistically significant differences are denoted with p -values, as determined by the t -test. Dependence of overall survival (OS) time on ulceration and VDR expression in the cytoplasm (C) and nuclei (D) of melanoma cells. Dependence of overall survival time on VDR expression in the nuclei (E) and cytoplasm (F) of melanoma cells in cases without ulceration. Representative VDR immunostaining in melanoma without (G) and with (H) ulceration. Arrows indicate nuclear VDR immunostaining, arrowhead indicates cytoplasmic VDR immunostaining; scale bar: 50 μ m.

In patients with melanoma, both systemic and local vitamin D₃ synthesis may also affect biology of malignant cells (31). Population-based study showed that a low serum level of 25(OH)D₃ was related to more advanced disease (overall stage and Breslow thickness), poor prognosis and higher mortality risk (32, 33). Population-based studies also revealed that VDR gene polymorphisms can alter susceptibility and prognosis of patients with melanoma (34).

TILs have been found in many types of human cancer (e.g. colon, ovarian, and laryngological cancer) (35-37).

Their presence is also related to better prognosis and longer OS. In melanomas, the presence of brisk TILs is considered a good prognostic factor. A weak immune response in patients with melanomas, determined as a lack of brisk TILs is considered a significant predictor of sentinel lymph node metastasis and shorter DFS (25, 26). As was mentioned above, immune responses can be modulated by calcitriol, and immune cells express VDR and enzymes activating vitamin D₃ (4, 9, 38). In our previous study, we observed a high expression of VDR in TILs and lymph node lymphocytes

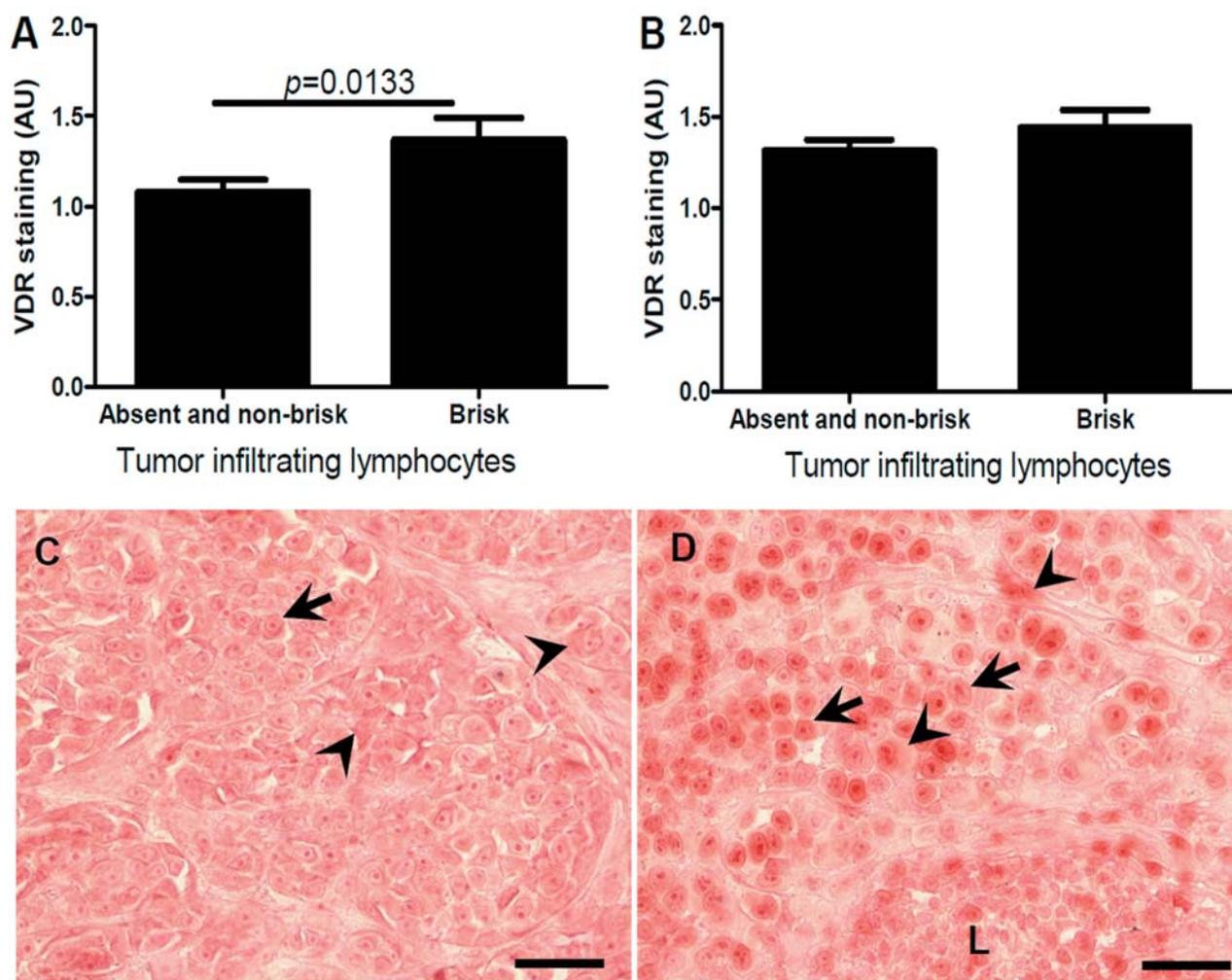


Figure 4. Mean cytoplasmic (A) and nuclear (B) expression of vitamin D receptor (VDR) in melanomas with absent or non-brisk tumor infiltrating lymphocytes (TILs). Statistically significant differences are denoted with p-values, as determined by the t-test. Representative VDR immunostaining in melanomas without (C) or with brisk (D) TILs. Arrows indicate nuclear VDR immunostaining, arrowhead indicates cytoplasmic VDR immunostaining, L marks lymphocytes; scale bar: 50 μm.

(22). In the present study, VDR expression in melanoma cells was positively-related to the presence of brisk TILs, suggesting that activation of vitamin D₃ by lymphocytes could negatively-affect proliferation of melanoma cells, resulting in better prognosis and outcome. Activation of the vitamin D₃-mediated signaling pathway could represent an additional mechanism of tumor-specific action of T-cells.

Another prognostic marker in patients with melanoma is ulceration, and its presence is a hallmark of more aggressive tumor biology, related to shorter OS (39). In our study, we found a significantly lower level of VDR expression both in the cytoplasm and in the nucleus in ulcerated melanomas in comparison to melanomas without ulceration. Accordingly, in nonulcerated melanomas, high expression of VDR was related to a longer OS.

In conclusion, we propose that increased VDR expression allows for prediction of a better prognosis, as demonstrated by its correlation with prognostic factors such as TIL presence, ulceration, overall stage and pTNM classification. Accordingly, loss of VDR should affect melanoma behavior, allowing it to evade host surveillance, and releasing the break on cell proliferation that would lead to progression of disease.

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References

- 1 MacKie RM, Hauschild A and Eggermont AM: Epidemiology of invasive cutaneous melanoma. *Ann Oncol* 20(Suppl 6): vi1-7, 2009.
- 2 Slominski A, Tobin DJ, Shibahara S and Wortsman J: Melanin pigmentation in mammalian skin and its hormonal regulation. *Physiol Rev* 84: 1155-1228, 2004.
- 3 Slominski A, Zmijewski MA and Pawelek J: L-Tyrosine and L-dihydroxyphenylalanine as hormone-like regulators of melanocyte functions. *Pigment Cell Melanoma Res* 25: 14-27, 2012.
- 4 Holick MF: Vitamin D deficiency. *N Engl J Med* 357: 266-281, 2007.
- 5 Bikle DD: Vitamin D: an ancient hormone. *Exp Dermatol* 20: 7-13, 2011.
- 6 Slominski AT, Zmijewski MA, Skobowiat C, Zbytek B, Slominski RM and Stekete JD: Sensing the environment: regulation of local and global homeostasis by the skin's neuroendocrine system. *Adv Anat Embryol Cell Biol* 212: v-115, 2012.
- 7 Holick MF: Sunlight, UV-radiation, vitamin D and skin cancer: How much sunlight do we need? *Adv Exp Med Biol* 624: 1-15, 2008.
- 8 Slominski AT, Kim TK, Shehabi HZ, Tang EK, Benson HA, Semak I, Lin Z, Yates CR, Wang J, Li W and Tuckey RC: *In vivo* production of novel vitamin D₂ hydroxy-derivatives by human placentas, epidermal keratinocytes, Caco-2 colon cells and the adrenal gland. *Molecular and cellular endocrinology* 383: 181-192, 2014.
- 9 Slominski AT, Kim TK, Li W, Yi AK, Postlethwaite A and Tuckey RC: The role of CYP11A1 in the production of vitamin D metabolites and their role in the regulation of epidermal functions. *J Steroid Biochem Mol Biol* 2013.
- 10 Slominski A, Janjetovic Z, Tuckey RC, Nguyen MN, Bhattacharya KG, Wang J, Li W, Jiao Y, Gu W, Brown M and Postlethwaite AE: 20S-Hydroxyvitamin D₃, noncalcemic product of CYP11A1 action on vitamin D₃, exhibits potent antifibrogenic activity *in vivo*. *J Clin Endocrinol Metab* 98: E298-303, 2013.
- 11 Slominski AT, Kim TK, Shehabi HZ, Semak I, Tang EK, Nguyen MN, Benson HA, Korik E, Janjetovic Z, Chen J, Yates CR, Postlethwaite A, Li W and Tuckey RC: *In vivo* evidence for a novel pathway of vitamin D(3) metabolism initiated by P450scc and modified by CYP27B1. *FASEB J* 26: 3901-3915, 2012.
- 12 Bikle DD: Vitamin D and immune function: understanding common pathways. *Curr Osteoporos Rep* 7: 58-63, 2009.
- 13 Reichrath J: Vitamin D and the skin: an ancient friend, revisited. *Exp Dermatol* 16: 618-625, 2007.
- 14 Chiang KC and Chen TC: The anticancer actions of vitamin D. *Anticancer Agents in Medicinal Chemistry* 13: 126-139, 2013.
- 15 Campbell FC, Xu H, El-Tanani M, Crowe P and Bingham V: The yin and yang of vitamin D receptor (VDR) signaling in neoplastic progression: operational networks and tissue-specific growth control. *Biochem Pharmacol* 79: 1-9, 2010.
- 16 Slominski A, T-K K, Takeda Y, Janjetovic Z, Brozyna AA, Skobowiate C, Wang J, Postlethwaite A, Li W, Tuckey R and Jetten A: ROR α and ROR γ are expressed in human skin and serve as receptors for endogenously produced noncalcemic 20-hydroxy- and 20,23-dihydroxy-vitamin D. *FASEB J* 2014 in press. Published online before print March 25, 2014, doi: 10.1096/fj.13-242040.
- 17 Slominski AT, Janjetovic Z, Fuller BE, Zmijewski MA, Tuckey RC, Nguyen MN, Sweatman T, Li W, Zjawiony J, Miller D, Chen TC, Lozanski G and Holick MF: Products of vitamin D₃ or 7-dehydrocholesterol metabolism by cytochrome P450scc show anti-leukemia effects, having low or absent calcemic activity. *PLoS One* 5: e9907, 2010.
- 18 Slominski AT, Kim TK, Janjetovic Z, Tuckey RC, Bieniek R, Yue J, Li W, Chen J, Nguyen MN, Tang EK, Miller D, Chen TC and Holick M: 20-Hydroxyvitamin D₂ is a noncalcemic analog of vitamin D with potent antiproliferative and prodifferentiation activities in normal and malignant cells. *Am J Physiol Cell Physiol* 300: C526-541, 2010.
- 19 Wang J, Slominski A, Tuckey RC, Janjetovic Z, Kulkarni A, Chen J, Postlethwaite AE, Miller D and Li W: 20-Hydroxyvitamin D(3) inhibits proliferation of cancer cells with high efficacy while being non-toxic. *Anticancer Res* 32: 739-746, 2012.
- 20 Slominski AT, Janjetovic Z, Kim TK, Wright AC, Grese LN, Riney SJ, Nguyen MN and Tuckey RC: Novel vitamin D hydroxyderivatives inhibit melanoma growth and show differential effects on normal melanocytes. *Anticancer Res* 32: 3733-3742, 2012.
- 21 Szyszka P, Zmijewski MA and Slominski AT: New vitamin D analogs as potential therapeutics in melanoma. *Expert Rev Anticancer Ther* 12: 585-599, 2012.
- 22 Brozyna AA, Jozwicki W, Janjetovic Z and Slominski AT: Expression of vitamin D receptor decreases during progression of pigmented skin lesions. *Hum Pathol* 42: 618-631, 2011.
- 23 Brozyna AA, Jozwicki W, Carlson JA and Slominski AT: Melanogenesis affects overall and disease-free survival in patients with stage III and IV melanoma. *Hum Pathol* 44: 2071-2074, 2013.
- 24 Brozyna AA, Jozwicki W, Janjetovic Z and Slominski AT: Expression of the vitamin D-activating enzyme 1 alpha-hydroxylase (CYP27B1) decreases during melanoma progression. *Hum Pathol* 44: 374-387, 2013.
- 25 Burton AL, Roach BA, Mays MP, Chen AF, Ginter BA, Vierling AM, Scoggins CR, Martin RC, Stromberg AJ, Hagendoorn L and McMasters KM: Prognostic significance of tumor infiltrating lymphocytes in melanoma. *Am Surg* 77: 188-192, 2011.
- 26 Mihm MC, Jr., Clemente CG and Cascinelli N: Tumor infiltrating lymphocytes in lymph node melanoma metastases: a histopathologic prognostic indicator and an expression of local immune response. *Lab Invest* 74: 43-47, 1996.
- 27 Janjetovic Z, Brozyna AA, Tuckey RC, Kim TK, Nguyen MN, Jozwicki W, Pfeffer SR, Pfeffer LM and Slominski AT: High basal NF- κ B activity in nonpigmented melanoma cells is associated with an enhanced sensitivity to vitamin D₃ derivatives. *Br J Cancer* 105: 1874-1884, 2011.
- 28 Colston K, Colston MJ and Feldman D: 1,25-Dihydroxyvitamin D₃ and malignant melanoma: the presence of receptors and inhibition of cell growth in culture. *Endocrinology* 108: 1083-1086, 1981.
- 29 Reichrath J, Rech M, Moeini M, Meese E, Tilgen W and Seifert M: *In vitro* comparison of the vitamin D endocrine system in 1,25(OH)₂D₃-responsive and -resistant melanoma cells. *Cancer Biol Ther* 6: 48-55, 2007.
- 30 Yudoh K, Matsuno H and Kimura T: 1 Alpha,25-dihydroxyvitamin D₃ inhibits *in vitro* invasiveness through the extracellular matrix and *in vivo* pulmonary metastasis of B16 mouse melanoma. *J Lab Clin Med* 133: 120-128, 1999.

- 31 Slominski AT and Carlson AJ: Melanoma resistance: bright future for academicians and challenge for patient advocates. *Mayo Clinic Proceedings* 89: 429-433, 2014.
- 32 Newton-Bishop JA, Beswick S, Randerson-Moor J, Chang YM, Affleck P, Elliott F, Chan M, Leake S, Karpavicius B, Haynes S, Kukulizch K, Whitaker L, Jackson S, Gerry E, Nolan C, Bertram C, Marsden J, Elder DE, Barrett JH and Bishop DT: Serum 25-hydroxyvitamin D3 levels are associated with Breslow thickness at presentation and survival from melanoma. *J Clin Oncol* 27: 5439-5444, 2009.
- 33 Nurnberg B, Graber S, Gartner B, Geisel J, Pfohler C, Schadendorf D, Tilgen W and Reichrath J: Reduced serum 25-hydroxyvitamin D levels in stage IV melanoma patients. *Anticancer Res* 29: 3669-3674, 2009.
- 34 Li C, Liu Z, Zhang Z, Strom SS, Gershenwald JE, Prieto VG, Lee JE, Ross MI, Mansfield PF, Cormier JN, Duvic M, Grimm EA and Wei Q: Genetic variants of the vitamin D receptor gene alter risk of cutaneous melanoma. *J Invest Dermatol* 127: 276-280, 2007.
- 35 Deschoolmeester V, Baay M, Van Marck E, Weyler J, Vermeulen P, Lardon F and Vermorken JB: Tumor infiltrating lymphocytes: an intriguing player in the survival of colorectal cancer patients. *BMC Immunol* 11: 19, 2010.
- 36 Sato E, Olson SH, Ahn J, Bundy B, Nishikawa H, Qian F, Jungbluth AA, Frosina D, Gnjjatic S, Ambrosone C, Kepner J, Odunsi T, Ritter G, Lele S, Chen YT, Ohtani H, Old LJ and Odunsi K: Intraepithelial CD8⁺ tumor-infiltrating lymphocytes and a high CD8⁺/regulatory T-cell ratio are associated with favorable prognosis in ovarian cancer. *Proc Natl Acad Sci USA* 102: 18538-18543, 2005.
- 37 Zhang L, Conejo-Garcia JR, Katsaros D, Gimotty PA, Massobrio M, Regnani G, Makrigiannakis A, Gray H, Schlienger K, Liebman MN, Rubin SC and Coukos G: Intratumoral T-cells, recurrence, and survival in epithelial ovarian cancer. *N Engl J Med* 348: 203-213, 2003.
- 38 Bikle DD: Vitamin D regulation of immune function. *Vitamins and hormones* 86: 1-21, 2011.
- 39 Balch CM, Wilkerson JA, Murad TM, Soong SJ, Ingalls AL and Maddox WA: The prognostic significance of ulceration of cutaneous melanoma. *Cancer* 45: 3012-3017, 1980.

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